

Synthesis of Jenamidines A₁/A₂

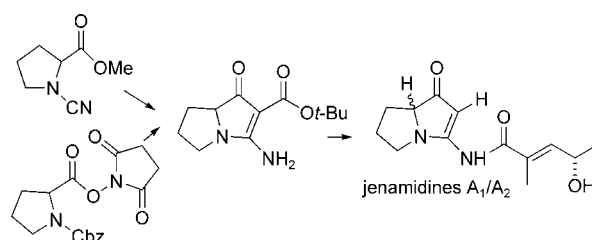
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Received August 4, 2005

ABSTRACT



Addition of the enolate of *tert*-butyl acetate to cyanamide methyl ester **17** followed by treatment with LHMDS afforded vinylogous urea **19** in 27% yield. Vinylogous urea **19** was also obtained from **37** and *tert*-butyl cyanoacetate in 50% yield. Acylation of **19** with acid chloride **31d**, followed by hydrolysis of the *tert*-butyl ester and decarboxylation with 9:1 CH₂Cl₂/TFA and very mild basic hydrolysis of the methoxyacetate ester, afforded jenamidines A₁/A₂ (**3**) in 45% yield. This first synthesis confirms our reassignment of the jenamidines A₁/A₂ structure.

Three bicyclic alkaloids, jenamidines A–C, were recently isolated from the culture broth of *Streptomyces* sp. (strain HKI0297).¹ Jenamidine A inhibited proliferation of the chronic myeloid leukemia cell line K-562 (GI₅₀ = 1.9 μg/mL). Structure **1** was originally proposed for jenamidine A (see Figure 1). We prepared model **2**, which underwent a facile retro-Mannich reaction and had spectral data quite different from jenamidine A, suggesting that structure **1** is not correct.² Reexamination of the spectral data of the natural product led to revised structures for the two diastereomers of jenamidines A₁/A₂ (**3**), the two diastereomers of jenamidines B₁/B₂ (**4**), and jenamidine C (**5**).² Bohemamine (**6**), whose structure was determined by X-ray crystallography in 1980,³ and the cell–cell adhesion inhibitor NP25302 (**7**),⁴ whose structure was reported very recently, have the same ring system as the revised structures of jenamidines **3**–**5**.

We next turned our attention to the preparation of jenamidines A₁/A₂ (**3**), which required the development of new methods for the preparation of the novel *N*-acyl

vinylogous urea in the right-hand ring. We initially explored the Pd-catalyzed coupling of triflate **8** with an amide since

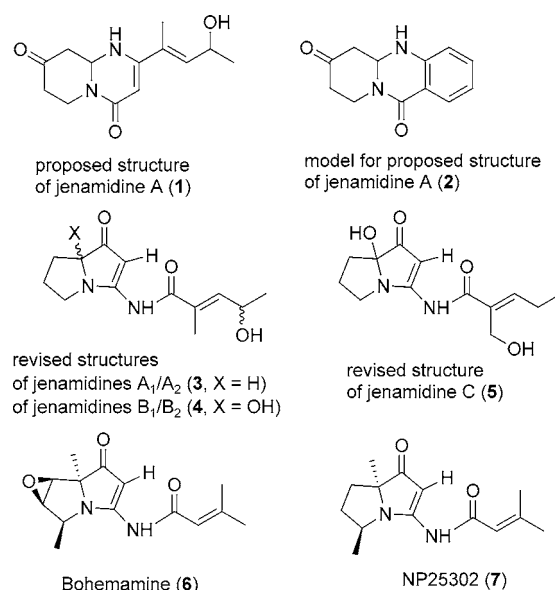


Figure 1. Structures of jenamidines and related natural products.

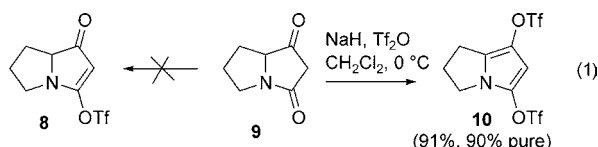
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(2) Snider, B. B.; Duvall, J. R.; Sattler, I.; Huang, X. *Tetrahedron Lett.* **2004**, *45*, 6725–6727.

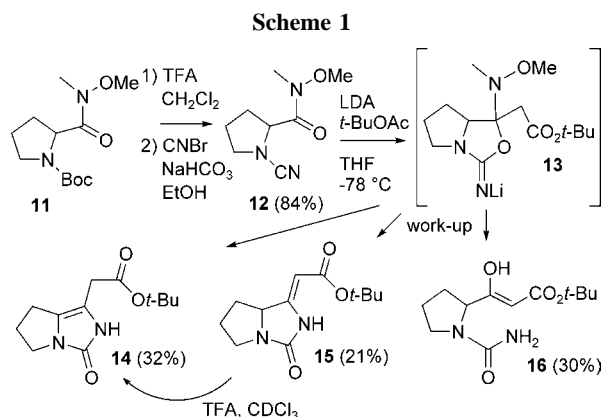
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a broadly applicable Pd-catalyzed amidation of enol triflates was recently reported.⁵ Unfortunately, reaction of known keto lactam **9**⁶ with NaH and Tf₂O gave only the unstable pyrrole bis triflate **10**. Use of excess NaH and Tf₂O gave crude (90% pure) **10** in 91% yield, which was isolated in pure form in only 17% yield (see eq 1). Although we were able to cleanly couple 2-methyl-2-butenamide⁷ with the enol triflate prepared from 5,5-dimethyl-1,3-cyclohexanedione, initial attempts at Pd-catalyzed couplings of amides with **10** were unsuccessful. Attempted preparation of vinylogous urea **33** (see Scheme 5) by reaction of keto lactam **9** with NH₃ led to complex mixtures.



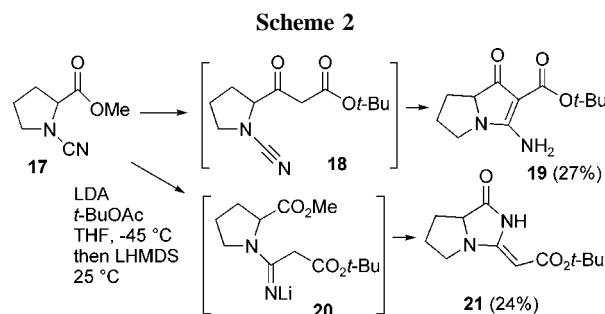
We then turned our attention to preparing the vinylogous urea by addition of an enolate to a cyanamide. Hydrolysis of the Boc group of Weinreb amide **11** and reaction of the liberated amine with CNBr and NaHCO₃ in EtOH afforded cyanamide **12** in 84% yield (see Scheme 1). Addition of the



lithium enolate of *tert*-butyl acetate to **12** provided 30% of the enol tautomer of urea β -keto ester **16**, 21% of imidazolidinone **15**, and 32% of imidazolone **14**. Presumably the lithium alkoxide of the initially formed tetrahedral intermediate adds to the cyanamide to give **13**. Workup affords urea keto ester **16**, which can undergo cyclodehydration to give **14** and **15**. Imidazolone **14** is the thermodynamic product since treating a solution of **15** in CDCl₃ with one drop of TFA cleanly isomerized **15** to **14**.

The Weinreb amide appeared to be a poor choice because the initially formed tetrahedral intermediate was stable,

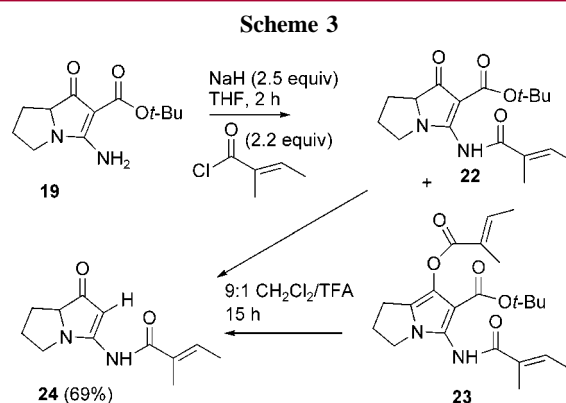
allowing the alkoxide to add to the cyanamide to form **13**. We thought that a simple ester might be a better choice because the tetrahedral intermediate should rapidly form the cyanamide keto ester **18**, which could then cyclize to form the desired product **19**. Fortunately, this proved to be the case. Cyanamide methyl ester **17**⁸ was added to a solution of the lithium enolate of *tert*-butyl acetate (2.3 equiv) in THF at -45 °C.⁹ The solution was stirred for 1 h at -45 °C, treated with 1.2 equiv of LHMDS in THF, and stirred at 25 °C for 2 h to give the desired product **19** (27%) (see Scheme 2). Byproduct **21** (24%) was formed by addition of



the enolate to the cyanamide to give **20**, which then cyclized to the methyl ester to form the alkylidene imidazolidinone **21**.¹⁰ The methyl ester of **17** is less electrophilic than the Weinreb amide of **12** so that the enolate added to both the methyl ester and the cyanamide.

Vinylogous urea **19** has the ring system of jenamidines A₁/A₂ with an additional carboxylic acid, which we hoped that we could remove by hydrolysis and decarboxylation either before or after the introduction of the side chain. Reaction of **19** with 9:1 CH₂Cl₂/TFA effected hydrolysis but did not provide the desired vinylogous urea **33** (see Scheme 5).

Acylation of **19** with 2.5 equiv of NaH and 2.2 equiv of tigloyl chloride for 2 h afforded a mixture of the desired amide **22** and the bis-acylated product pyrrole **23** (see Scheme 3). Treatment of the crude mixture with 9:1 CH₂Cl₂/TFA for 15 h effected hydrolysis of the *tert*-butyl esters of **22** and **23** and the enol ester of **23** and decarboxy-



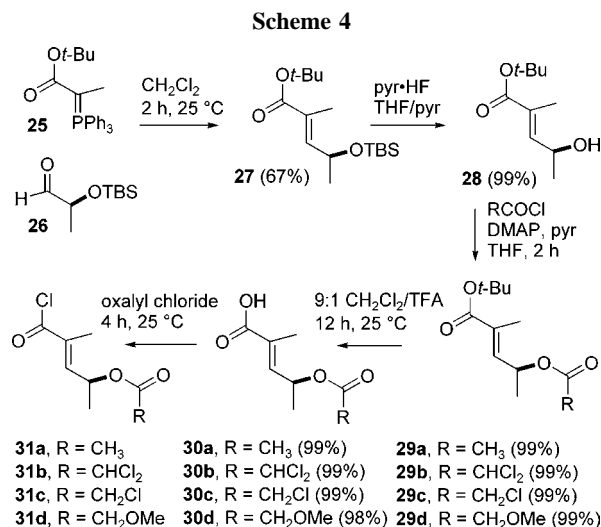
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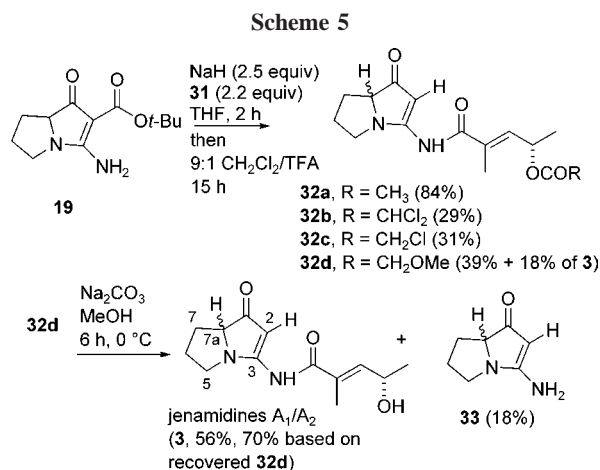
lation to afford jenamidines A₁/A₂ model **24** in 69% overall yield. The spectral data of the ring portion of **24** correspond very closely to those of the natural product, supporting the assignment of **3** as the revised structure of jenamidines A₁/A₂.

The side chain was then prepared by a modification of Adam's procedure for the ethyl ester.¹¹ Reaction of ylide **25**¹² with aldehyde **26**¹³ in CH₂Cl₂ for 2 h provided ester **27** in 67% yield (see Scheme 4). Deprotection with pyr·HF gave



hydroxy ester **28** in 99% yield. Initially, we chose to protect the side chain alcohol as an acetate ester. Reaction of **28** with AcCl, DMAP and pyridine in THF gave **29a** in 99% yield, which was deprotected with 9:1 CH₂Cl₂/TFA to give acetoxy acid **30a** in 99% yield. Stirring **30a** in oxalyl chloride gave crude acid chloride **31a**, which was used without purification.

Reaction of **19** with NaH and **31a** followed by hydrolysis and decarboxylation with 9:1 CH₂Cl₂/TFA as described above for the preparation of **24** afforded jenamidines A₁/A₂ acetate (**32a**) in 84% yield (see Scheme 5). Unfortunately, we were unable to cleave the acetate protecting group of **32a** without also cleaving the side chain amide to give a complex mixture containing some **33**. Since the nitrogen of the amide of **32a** is part of a vinylogous urea, the amide is a vinylogous acyl urea and is therefore easily cleaved under basic conditions. We considered using an acid-labile protect-



ing group for the side chain alcohol that would be cleaved by the 9:1 CH₂Cl₂/TFA used for hydrolysis of the *tert*-butyl ester. Unfortunately, such a protecting group would not be compatible with acid chloride **31**, and we were unable to cleanly acylate **19** with mixed anhydrides.

We then examined more base labile ester protecting groups.¹⁴ Dichloroacetate acid chloride **31b** was prepared analogously, but reaction with **19** afforded **32b** in only 29% yield. Fortunately, hydrolysis of **32b** with NaHCO₃ in MeOH for 30 min at 25 °C gave jenamidines A₁/A₂ (**3**) in 71% yield. Reaction of chloroacetate **31c** with **19** afforded **32c** in a still unacceptable 31% yield, which could also be cleaved by NaHCO₃ in MeOH for 1 h at 25 °C to give jenamidines A₁/A₂ (**3**) cleanly.

The best compromise was the methoxyacetate protecting group. Acid chloride **31d** was prepared in high yield from hydroxy ester **28**. Coupling of **31d** with **19**, hydrolysis of the *tert*-butyl ester, decarboxylation with 9:1 CH₂Cl₂/TFA, and flash chromatography on silica gel gave **32d** in 39% yield and jenamidines A₁/A₂ (**3**) in 18% yield. Partial cleavage of the methoxyacetate occurs on chromatography. Hydrolysis of **32d** with Na₂CO₃ in MeOH for 6 h at 0 °C provided **3** in 56% yield (70% based on recovered **32d**) and 18% of **33** resulting from cleavage of the amide. Hydrolysis of **32d** with NaHCO₃ in MeOH for 20 h at 25 °C afforded only **33** indicating the sensitivity of the amide side chain to basic hydrolysis. The most efficient procedure involved hydrolysis of crude **32d** with Na₂CO₃ in MeOH for 24 h at 0 °C to give jenamidines A₁/A₂ (**3**) in 45% overall yield from **19** and **32d** in 11% overall yield from **19**.

The spectral data of synthetic **3** are identical to those of the natural product, which is also an approximately 1:1 mixture of diastereomers. Even though **19** was prepared from (*S*)-proline and aldehyde **26** was prepared from (*S*)-lactic acid, we obtained **3** as a mixture of diastereomers. The ring fusion hydrogen is readily epimerized and this stereocenter is lost in the formation of the bis acylated intermediate

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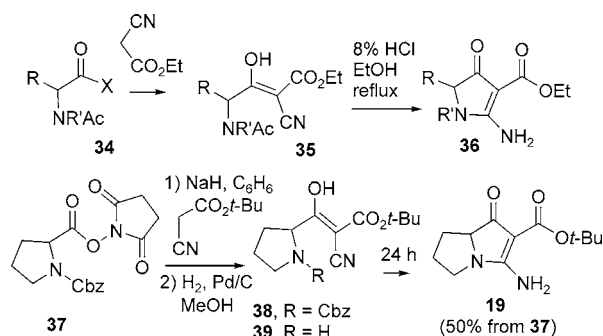
(14) Kocięński, P. J. *Protecting Groups*, 3rd ed.; Georg Thieme Verlag: Stuttgart, 2005; pp 333–337.

analogous to **23**, which will give a mixture of diastereomers on hydrolysis. In the proton NMR spectrum of **3** in CD₃OD, the ring fusion hydrogen, H-7a, integrates for only ~0.5, suggesting that partial deuterium exchange has occurred. C-2 and C-7 absorb as four peaks since a separate peak is observed for the H-7a and D-7a isomer of each diastereomer.¹⁵ H-2 slowly exchanges with CD₃OD over several hours as was noted for the natural product.² The optical rotation of synthetic **3**, [α]_D 4.2, is very similar to that of the natural product, [α]_D 6.8.¹ Therefore, natural jenamidines A₁/A₂ (**3**) could also be a mixture of isomers at the ring fusion and the (*S*)-isomer on the side chain. However, since both rotations are for mixtures of isomers, it is also possible that the natural product is a mixture of isomers on the side chain.

The three-step sequence from vinylogous urea **19** and acid chloride **31d** to jenamidines A₁/A₂ (**3**) proceeded in acceptable yield, given the instability of the amide linkage. The one-pot preparation of **19** from cyanamide **17** provided adequate quantities of material, but the 27% yield left room for improvement. Coupling of various *N*-acetyl amino acid derivatives **34** with ethyl cyanoacetate has been reported to give **35**, which cyclized on treatment with 8% HCl in EtOH at reflux to provide **36** in 18–51% overall yield (see Scheme 6).¹⁶ The reported spectral data of **36** are comparable to those of **19**. We examined variations of this procedure because the acid-catalyzed cyclization used to convert **35** to **36** is not compatible with the *tert*-butyl ester of **19**.

Reaction of Cbz-proline *N*-hydroxysuccinimide ester (**37**) with *tert*-butyl cyanoacetate and NaH in benzene for 3 h

Scheme 6



gave crude **38**, which was hydrogenated (1 atm) over 10% Pd/C in MeOH for 2 h to give crude **39** with a very complex NMR spectrum. Fortunately, crude **39** cyclized on standing for 1 d to give **19** in 50% overall yield from **37**. Using this sequence, which has not been fully optimized, jenamidines A₁/A₂ (**3**) are now available in 23% overall yield.

In conclusion, addition of the enolate of *tert*-butyl acetate to cyanamide methyl ester **17** followed by treatment with LHMDS afforded vinylogous urea **19** in 27% yield. Vinylogous urea **19** can be obtained more easily from **37** and *tert*-butyl cyanoacetate in 50% yield. Acylation of **19** with acid chloride **31d**, followed by hydrolysis of the *tert*-butyl ester and decarboxylation with 9:1 CH₂Cl₂/TFA and very mild basic hydrolysis of the methoxyacetate ester afforded jenamidines A₁/A₂ (**3**) in 45% yield. This first synthesis confirms our reassignment of the jenamidines A₁/A₂ structure. Extension of this approach to the syntheses of jenamidines B₁/B₂, jenamidine C, and NP25302 is currently in progress.

Acknowledgment. We thank the NIH (GM50151) for generous financial support. We thank Prof. Isabel Sattler for helpful discussions.

Supporting Information Available: Full experimental details and copies of ¹H and ¹³C NMR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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